





Crossing paths with Notch in the hyper-network Gregory D Hurlbut^{1,2}, Mark W Kankel¹, Robert J Lake¹ and Spyros Artavanis-Tsakonas^{1,3}

The development of complex and diverse metazoan morphologies is coordinated by a surprisingly small number of evolutionarily conserved signaling mechanisms. These signals can act in parallel but often appear to function as an integrated hyper-network. The nodes defining this complex molecular circuitry are poorly understood, but the biological significance of pathway cross-talk is profound. The importance of such large-scale signal integration is exemplified by Notch and its ability to cross-talk with all the major pathways to influence cell differentiation, proliferation, survival and migration. The Notch pathway is, thus, a useful paradigm to illustrate the complexity of pathway cross-talk: its pervasiveness, context dependency, and importance in development and disease.

Addresses

¹ Department of Cell Biology, Harvard Medical School, Massachusetts General Hospital Cancer Center, Charlestown, Massachusetts, USA ² Faculté des Sciences d'Orsay, Université de Paris-Sud 11, Paris, France

³Collège de France, Paris, France

Corresponding author: Artavanis-Tsakonas, Spyros (tsakonas@helix.mgh.harvard.edu)

Current Opinion in Cell Biology 2007, 19:166-175

This review comes from a themed issue on Cell regulation Edited by Carl-Henrik Heldin and Peter ten Dijke

Available online 20th February 2007

0955-0674/\$ - see front matter © 2006 Elsevier Ltd. All rights reserved.

DOI 10.1016/j.ceb.2007.02.012

Introduction

Metazoans rely on a handful of core signaling mechanisms to guide a wide range of developmental processes, from the earliest specification events to organogenesis. Key among these are the Hedgehog (Hh), Janus kinase/signal transducers and activators of transcription (Jak/STAT), nuclear receptor, receptor tyrosine kinase (RTK), transforming growth factor- β /Decapentaplegic (TGF- β /Dpp), Wnt/Wingless (Wnt), and Notch (N) pathways [1–4]. Together, these highly conserved pathways create a signaling backbone supporting all stages of metazoan development [1–4]. It is remarkable that metazoan species, despite being constrained to this shared signaling framework, have managed to evolve into species of vastly diverse body plans [5^{••}]. To achieve the morphological complexity that is characteristic of metazoans, these core signaling pathways must integrate to form a larger, complex signaling system, which we term the hyper-network. However, comprehensive knowledge of this network, the nodes that define it and its emergent properties is lacking. Studying how these highly pleiotropic pathways are interlinked is essential to understanding development and evolution and, consequently, defines a fundamental problem in biology with obvious implications for disease.

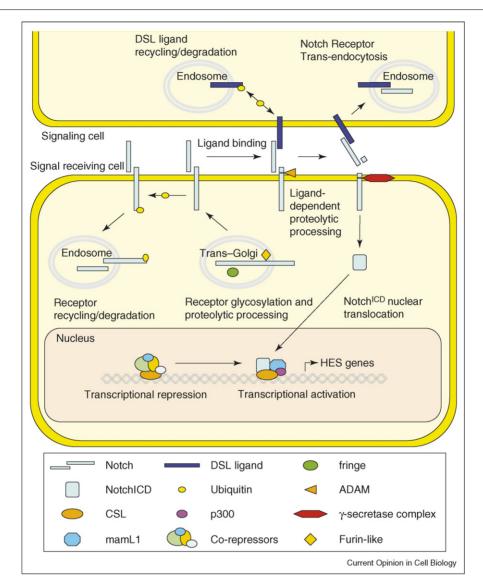
Given the pleiotropy of Notch signaling, its importance to development and disease $[6^{\bullet\bullet},7-10]$, and its ability to integrate with all major pathways (see below and Figure 1), in this review we focus on Notch signal integration, or 'cross-talk'. We aim to provide a brief perspective on this signaling hyper-network and illustrate the importance of cross-talk, its pervasiveness, and its capacity to generate complexity during development.

Metazoans share common functional and mechanistic aspects of Notch signaling, which have been outlined in several recent reviews [6^{••},7–10]. Notch signaling involves receptor activation by a membrane-bound Delta/Serrate/Lag-2 (DSL) ligand, leading to proteolytic processing of the receptor (Figure 1). This releases the central signaling molecule, the Notch intracellular domain (N^{ICD}), which undergoes nuclear translocation and association with a CBF1/Su(H)/Lag-1 (CSL) family transcription factor, promoting expression of E(spl)/HES family and other target genes [11]. Ultimately, Notch signaling affects cell-fate specification, proliferation, apoptosis and migration. Aberrant Notch signaling has been associated with pathogenic conditions including carcinogenesis. Very few studies have been specifically designed to address pathway cross-talk; however, numerous links between Notch and other signaling pathways have emerged (Figure 2). Undoubtedly, Notch cross-talk is pervasive in development and contributes to the astounding spectrum of Notch function (Table 1).

Cell-fate specification

Cross-talk has an important and prevalent role in cell-fate specification. As exemplified by Notch/RTK integration, the influence of cross-talk on cell fate appears to be complex, and, in different contexts, integration can have agonistic or antagonistic effects. Antagonism predominates during *C. elegans* vulval development and in some aspects of *Drosophila* photoreceptor development. In both these cases, Notch opposes RTK-mediated induction of differentiation [12–15]. In *Drosophila*, such antagonism is

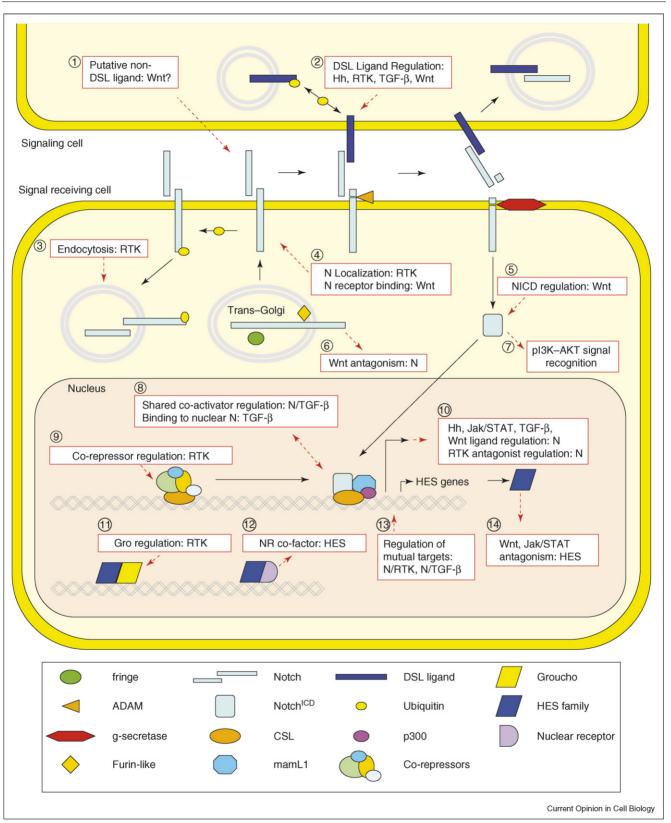




A current overview of Notch signal transduction. The Notch pathway mediates regulation of a diverse array of cell-fate decisions through juxtacrine signaling. Notch receptors are composed of an extracellular domain (N^{ECD}) containing numerous Ca²⁺-binding EGF-like repeats, a small transmembrane region (NTM), and an intracellular domain (N^{icd}) that can act as a nuclear effector. In the ER and Golgi, N^{ECD} is modified by a series of glycosylation events mediated by Fringe and other glycosyltransferases. In the *trans*-Golgi, Notch undergoes proteolysis by a furin-like convertase, generating a glycosylated, presumably divalent-cation-stabilized Notch heterodimer. In *Drosophila*, two DSL ligands, *Delta* and *Serrate*, activate signaling. In mammals, ligands include members of the Delta-like (DLL1, DLL3, DLL4) and Jagged (JAG1, JAG2) families. Upon ligand binding, N^{ECD} is removed through cleavage by an ADAM metalloprotease, TNF- α converting enzyme (TACE). N^{ECD} remains bound to the ligand, and both proteins may be endocytosed by the ligand presenting cell. The physical force generated by this ligand/N^{ECD} internalization may be required for receptor cleavage by TACE, and further receptor processing required for signaling. Upon cleavage by TACE, constitutive cleavage events mediated by the γ -secretase complex release N^{ICD}. The N^{ICD} translocates to the nucleus and associates with a *CBF1/Su(H)/Lag-1* (*CSL*) family transcriptional regulator. In the absence of N^{ICD}, CSL family proteins are part of a repressor complex. Upon Notch binding, co-repressors are exchanged for co-activators, including Mastermind and p300, leading to the activation of target genes, including HES family members. Both the Notch receptor and its ligands can undergo ubiquitin-regulated internalization and degradation. We note that several aspects of this overview reflect a working hypothesis and some aspects are not necessarily rigorously proven. This is especially true for aspects of receptor and ligand trafficking and the

crucial to specification in numerous other contexts [14– 17]. Though, canonically, Notch signaling maintains an undifferentiated cell fate, in many contexts Notch is actually required to direct specification events, and here the effects of cross-talk may also be important. For example, in the *Drosophila* eye, in addition to the pathway antagonism observed in photoreceptor development, Notch and RTK act cooperatively to specify accessory





For an overview of Notch signaling see Figure 1. Nodes of integration are indicated by text in numbered red boxes and references for the data in each is listed next to its corresponding number below. Some of the indicated modes of cross-talk, including the role of wingless as a

cells [14,18,19]. Such variable effects of Notch pathway cross-talk on cell-fate specification and its context dependency seem to be the rule rather than the exception.

Like Notch/RTK cross-talk, the effects of signal integration with Wnt on cell fate vary with context. During vertebrate osteoblastogenesis, ST-2 stromal cells differentiate as osteoblasts when treated with Wnt3a. In the presence of ectopic Notch1^{ICD}, however, these cells undergo adipogenesis. Thus, in this context, the effect of Notch on Wnt signaling is antagonistic [20[•]]. In contrast, pathway synergy is observed in the adult mouse epidermis, where Notch and Wnt/β-catenin cooperate to maintain postnatal hair growth. If either signal is blocked, hair follicles convert into cysts of interfollicular epidermis, while simultaneous activation of both pathways increases induction of ectopic hair follicles [21]. Osteoblastogenesis is also influenced by Notch/TGF-B cross-talk, where cooperation between Notch1 and the TGF-B ligand BMP-2 promotes differentiation [22]. During myogenesis, however, Notch and BMP-4 synergize to inhibit differentiation [23]. The opposing influence of cross-talk on cell fate in different contexts thus appears to be a common theme, suggesting that flexible integration is an essential feature of the hyper-network.

Proliferation and apoptosis

Notch, classically associated with cell differentiation, has also been shown to direct cells into proliferative or apoptotic states. Interestingly, Notch has both cellautonomous and non-cell-autonomous effects on mitotic activity, which in different contexts it can either promote or suppress. Though many aspects of Notch signaling in proliferation and apoptosis remain poorly understood, its potential to link these events with differentiation may be of particular relevance to dysproliferative states, including cancer. Again, developmental context appears to dictate how Notch activation affects the cell cycle. The logic of integration appears complex, as Notch signals can have either oncogenic or tumor-suppressive effects in tumors of the same type (see below). As with differentiation, this complexity may depend upon cross-talk.

To date, three distinct signaling pathways, Jak/STAT, RTK and Wnt, have been shown to affect proliferation, in part through integration with Notch. At the dorsoventral boundary (DV) of the *Drosophila* eye primordium, where Notch activation plays a crucial role regulating eye growth and patterning [24[•]], ectopic Notch activation induces dramatic proliferation through a non-cell-autonomous

mechanism that involves activation of Jak/STAT signaling through its ligand *unpaired* [24°-26°]. In the *Drosophila* wing disc, where multiple signals influence growth during development [27], the Notch and RTK pathways can synergize to promote dramatic overgrowth (Hurlbut and Artavanis-Tsakonas, unpublished). Cross-talk with Wnt/wingless also influences wing growth. Notch/Wnt integration induces proliferation in cells of the wing pouch and hinge [27] but cell-cycle arrest in those that form the wing margin sensory organs [28]. As Notch/Wnt integration regulates early precursor cell proliferation in mouse intestinal crypts [29°,30] the importance of such cross-talk in proliferation appears to be functionally, if not mechanistically, conserved.

Notch signals have been shown to influence cell death in both vertebrates and invertebrates. The influence of cross-talk on this process is evident in the *Drosophila* eye, where, during late pupal development, Notch provides a pro-apoptotic cue that opposes RTK-dependent survival [31]. Demonstrating the significance of crosstalk, in the absence of RTK signaling through the EGFR receptor, Notch is no longer required for apoptosis. By contrast, Notch and Wnt/wingless act together to promote apoptosis during earlier stages, where signaling through EGFR appears not to be involved [32]. Highlighting the importance of context, wingless opposes a Notch proapoptotic signal in the *Drosophila* wing [27].

Stem cells

Stem cell maintenance, crucial to regeneration, requires signaling. Given the potent ability of Notch to influence cellular differentiation, it is not surprising that Notch signaling has emerged as an important regulator of stem cells of the mammary gland, eye, skin, nervous system, bone marrow, stroma, gastrointestinal (GI) tract and ovary [33]. Often, Notch integration with other signaling pathways plays an essential role, with Notch/Wnt cross-talk being of particular importance. In both the GI tract [34] and early hematopoiesis [35^{••}], Notch and Wnt influence stem/precursor cell maintenance and, perhaps, proliferative potential, through their cooperative effect. In some contexts, stem cell survival may depend on Notch integration with multiple pathways. In murine somatic and human embryonic stem cells, Notch signaling activates the pI3K/Akt/mTOR pathway, leading to specific phosphorylation of the Jak/STAT mediator STAT3 at serine residue 727, which induces expression of target genes including Sonic Hedgehog (Shh), and promotes stem cell survival. This positive survival signal is opposed by

(Figure 2 Legend Continued) Notch ligand, are currently controversial. Dashed arrows provide the direction of influence. Arrows pointing to Notch pathway components indicate that the pathway(s) shown has an effect on Notch signaling, Box 2 [18,60,74–76], Box 3 [77*,78], Box 4 [77*,78–80], Box 5 [79], Box 9 [76], Box 13 [18,23,70,81–85]. Those pointing away indicate an effect of Notch on the pathway(s) listed, Box 6 [86], Box 7 [36**,87], Box 10 [24*,36**,66,73,88**,89], Box 14 [20*]. Double headed arrows, Box 1 [90], Box 8 [81,91,92], indicate mutual influence specific to the component or process listed. Also shown in the nucleus, is the influence of RTK signaling on the HES co-repressor Groucho, Box 11 [93*], and the influence of Notch signaling on the Nuclear Receptor pathway (NR) by Notch targets of the HES family, which can serve as Nuclear Receptor co-factors, Box 12 [94].

Table 1 The biological contexts of Notch cross-talk.				
Hh	D. melanogaster	Eye	Cooperate in eye development	[74]
Ηh	D. rerio	Vasculature	Artery/vein cell specification (with VEGF)	[95]
Ηh	H. sapiens	Meduloblastoma	Promote tumor growth and survival (with Wnt)	[59,60]
Jak/STAT	D. melanogaster	Eye	Cooperatively promote growth	[24°-26°]
Jak/STAT	D. melanogaster	Foregut	Cooperatively govern development	[96]
lak/STAT	D. melanogaster	Oogenesis	Specify stalk and pre-polar cell fate	[97–99]
lak/STAT	H. sapiens, M. musculus	Embryonic and somatic stem cells	Stem cell survival	[36**]
Jak/STAT	M. musculus	Central nervous system	Cooperate in astrocyte differentiation	[100 °]
lak/STAT	M. musculus	Neural stem cells	Specify stem cell survival versus differentiation	[36**]
RTK	C. elegans	Vulva	Specify vulval cell fates	[13]
RTK	D. melanogaster	Chordotonal organ	SOP fate specification	[17]
RTK	D. melanogaster	Eye	Photoreceptor and accessory cell specification	[14]
RTK	D. melanogaster	Eye/Antenal disc	Specify eye versus antennal identity	[101]
RTK	D. melanogaster	Mesoderm	Specify muscle cell subtypes	[18]
RTK	D. melanogaster	Notum Bristles	SMC fate specification	[16]
RTK	D. melanogaster	Trachea	Specify fusion cell fate	[64]
RTK	D. melanogaster	Wing	Specify wing vein versus intervein	[102]
RTK	D. melanogaster	Wing	Cooperatively promote growth	Unpublished
RTK	D. rerio	Vasculature	Artery/vein cell specification (with Shh)	[95]
RTK	H. sapiens	Cell migration	Notch blocks Ras induced migration in culture	[56]
RTK	H. sapiens	Oncogenesis	Act cooperatively in transformation or growth	[53,55]
RTK	H. sapiens	Thymocyte development	T-Cell lineage specification (Notch and TCR)	[103]
RTK		Tumor angiogenesis	Cooperatively promote angiogenesis	
RTK	H. sapiens M. musculus	Embryonic segmentation		[104]
RTK		, ,	Cooperatively regulate segmentation clock	[105]
RTK	M. musculus	Spinal cord development	Maintenance of the caudal neural plate	[106]
	M. musculus	Neurogenesis	Promote radial glial identity	[107]
RTK	M. musculus	Oncogenesis	Cooperation or antagonism	[54,56,57]
RTK	M. musculus	Pancreatic tumorigenesis	Cooperate in pathogenesis	[58]
ΓGF-β	D. melanogaster	Germline stem cells	Maintenance of stem cells and stem cell niche	[37]
ΓGF-β	D. melanogaster	Trachea	Specify fusion cell fate	[66]
ΓGF-β	D. rerio	Cardiac development	Promote Epithelial-to-mesenchymal transition	[108]
ΓGF-β	M. musculus	Embryonic endothelial cells	Regulation of migration	[70]
ΓGF-β	M. musculus	Endothelial migration	Notch blocks TGF- β induced migration in culture	[70]
ΓGF-β	M. musculus	Muscle	Cooperatively inhibit myogenic differentiation	[23]
ΓGF-β	M. musculus	Neurogenesis	Cooperatively inhibit neuroepithelial differentiation	
ΓGF-β	M. musculus	Osteoblastogenesis	Cooperatively promote differentiation	[22]
ΓGF-β	M. musculus	Prostate gland	Regulation of branching morphogenesis	[68]
ΓGF-β	M. musculus	Prostate gland	Regulates epithelial bud formation	[68]
Nnt	D. melanogaster	Epidermis	Interact genetically in this context	[109,110]
Nnt	D. melanogaster	Trachea	Specification of fusion cell fate	[67]
Vnt	D. melanogaster	Wing	Cooperatively regulate development	[80,86,109,111-11
Vnt	D. melanogaster	Wing margin sensory organs	Cooperatively induce cell cycle arrest	[28]
Vnt	D. melanogaster	Wing Pouch and Hinge	Cooperatively promote growth	[27,115]
Vnt	H. sapiens	Mammary duct morphogenesis	Antagonistically regulate branching	[69]
Vnt	H. sapiens	Meduloblastoma	Promote tumor growth and survival (with Hh)	[60]
Nnt	H. sapiens	Melanoma	Promote tumor growth and metastasis	[61]
Vnt	M. musculus	Epidermis	Cooperate in postnatal hair follicle induction	[21]
Wnt	M. musculus	Gastrointestinal tract	Cooperatively regulate stem cell proliferation	[30,34]
Wnt	M. musculus	Hematopoietic stem cells	Stem cell maintenance	[35**]
Wnt	M. musculus	Osteoblastogenesis	Specify cell fate	[20 [•]]
Wnt	M. musculus	Somites	Cooperate in somitogenesis	[75]
	m. museulus	Connico	ooporato in oonnogonoolo	[, 9]

signaling through Jak/p38-MAPK, downstream of ciliary neurotrophic factor (CNTF), which counters Notchresponsive STAT3 phosphorylation at serine 727. This signal instead promotes STAT3 phosphorylation at tyrosine residue 705 and, ultimately, cell differentiation [36^{••}]. In the *Drosophila* ovary, maintenance of germline stem cells (GSCs) requires a Notch/TGF- β signal feedback loop. Loss of either signal leads to GSC differentiation and exit from the stem-cell niche $[37^{\circ}]$. Dysregulation of the stem cell self-renewal process, which involves regulation of apoptosis, proliferation and differentiation, may be a significant event in the genesis of certain cancers. Indeed, it is speculated that blocking this process may be a useful therapy [38]. This might be achieved by targeting the nodal points of signal integration, thus modulating the effects of cross-talk.

Oncogenesis

In normal tissues, proliferation, differentiation and apoptosis exist in delicate balance, and it is the disturbance of such homeostasis that commonly underlies oncogenesis. Given the fundamental importance of signaling in the regulation of these processes, it is not remarkable that disruptions in all major signaling pathways, including Notch, have been associated with oncogenesis [39–49]. Notch, initially linked to cancer through its frequent mutation in T-Cell acute lymphoblastic leukemia [50,51], can, in some tumor-types, integrate with other pathways to affect the course of oncogenesis profoundly. However, the observed effects of cross-talk are often complex and context-dependent. For example, Notch signaling has been shown to either enhance [52-55] or suppress [56,57] the transformation and proliferation of tumors in which RTK is activated. Such inconsistencies are likely to reflect differences in the underlying signal integration in these tumors or associated contexts. In pancreatic tumorigenesis, EGFR signaling may act, in part, to induce Notch activation. Here, cooperation between these pathways may be important to pathogenesis [58]. In human and murine medulloblastoma, Notch and Shh synergize to promote tumor proliferation and survival [59]. In fact, in medulloblastomas, Shhdependent tumor growth and survival involves synergy with both Notch and Wnt [60]. Notch/Wnt cross-talk has also been suggested to be of importance to melanoma. Activation of Notch1 enhances primary melanoma cell growth and the potential for metastasis through β -catenin upregulation [61]. It is clear that manipulating Notch activity can modify cell fate, and, as first speculated over a decade ago, this capacity makes Notch a therapeutic target of potentially great significance [62]. As evidenced by the different impacts of Notch/RTK integration on tumorigenesis, the effects of Notch manipulation are difficult to predict a priori. As a result, a substantial knowledge of the signaling hyper-network underlying a targeted tumor may be necessary for optimal therapy.

Branching morphogenesis/migration

Networks of branched, tube-like structures, found in metazoan organs of numerous types, are formed through precise regulation of cell differentiation, proliferation, apoptosis, adhesion and migration. Notch is among the many signals crucial to branching morphogenesis, and here cross-talk has also been documented to be important. During Drosophila tracheal development, cross-talk between Notch and the Wnt/wingless, TGF- β /Dpp and RTK/FGFR pathways generates branch patterning through the cooperative specification of cell fate [63-67]. In vertebrates, Notch cross-talk functions in epithelial bud formation and branching of the developing prostate gland [68]. Specifically, Notch/TGF-B antagonism helps regulate prostate branching, while TGF-B signaling, activated by the BMP-7 ligand, limits Notch activation, and the epithelial bud formation it promotes, to subsets of cells within the urogenital epithelium. Cross-talk is also evident in the developing mouse mammary gland, where Notch4-mediated inhibition of branching is overcome by Wnt1 activation [69].

The influence of cross-talk on cell migration has been observed in MDCK and MDA-MB-435 cells, where expression of constitutively active Notch4 blocks hepatocyte growth factor (HGF)-induced Ras activation and cell migration [56]. Additionally, when murine embryonic endothelial cells are in contact, ligand-dependent Notch signaling is activated, and BMP/TGF- β -mediated migration is inhibited [70]. This mechanism linking migration to contact through cross-talk may be of crucial importance both in development and in the pathology of numerous diseases.

Conclusions

The pleiotropy observed for Notch signaling during development is in large part dependent on the ability of context to influence its activity. The basic features of Notch signaling may have emerged by the Precambrian era [71,72] and, as new metazoan species evolved, Notch signaling seems to have retained a central role in development: coupling the fate choices of adjacent cells. However, the mechanisms regulating the Notch signal in different developmental contexts did not necessarily remain invariant during evolution. During development, Notch activity is modulated through the regulation of ligand availability, Notch pathway component transcription and trafficking, and post-translational modification of both the receptor and its ligands. The capacity for modulation at multiple levels may allow other signals and additional context-dependent factors to converge with Notch at numerous points. Supporting this, known mechanisms of signal integration involve nodal points at each step of Notch signal transduction (Figure 2).

It is important to note that cross-talk does not require synchronous signals, as sequential signals can also integrate. Sequential integration can affect progenitor cells that are maintained by Notch and specified by subsequent signals. The mutual influence that pathways can exert on each other through the regulation of ligands is another example of sequential integration. Here, the effects of cross-talk can be non-cell-autonomous. In different contexts, the output of Notch signaling includes the Hh, Jak/STAT, TGF-B/Dpp and Wnt pathway ligands [24[•],36^{••},66,73]. Reciprocally, Notch ligands are an output of Hh, RTK, TGF-B/Dpp and Wnt/wingless signaling [18,60,74-76]. This suggests that feedback loops represent important interlinking mechanisms that help turn separate signals into a network. Though soluble Notch ligands have been observed in C. elegans, it is noteworthy that Notch signaling is unique in its general requirement for direct membrane-membrane contact. Integration with Notch might, therefore, be a useful method for pathways activated by diffusible ligands to gain spatial resolution. As such, Notch may play a prominent role in bridging major pathways.

Through either a direct interface between pathways or shared target sets, signal integration might allow a simple interconnected system to generate an extraordinarily diverse output. The dramatic range of morphologies that these few signals can create supports this notion. Studying its effects on the transcriptional output of integrated signals may be a useful approach to understanding crosstalk. Extrapolating from the Notch cross-talk paradigm, several features of the signaling hyper-network can be inferred. First, signaling pathways are remarkably interlinked and can integrate at the cellular and multicellular level through multiple mechanisms. Second, cross-talk is of broad importance, impacting numerous pathway functions, and its dysregulation appears profoundly important to cancer, and potentially to other diseases. Third, cross-talk is flexible, differing in consequence in different contexts. The unknown source(s) providing this specificity are of fundamental importance. As multiple mechanisms interlink diverse signaling pathways with Notch, it is possible that the flexible nature of cross-talk depends upon the primary mechanism employed in each context, but a wide range of factors may also be involved, including additional cross-talk.

Acknowledgements

We sincerely apologize to those whose work could not be cited due to space constraints. Many thanks to James Walker and Doug Dimlich for improving the figures through their helpful suggestions. We are also indebted to Angeliki Louvi for her critical reading of the manuscript. The laboratory of S A-T is funded by the National Institutes of Health, the Ellison Foundation and the Spinal Muscular Atrophy Foundation.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- Barolo S, Posakony JW: Three habits of highly effective 1. signaling pathways: principles of transcriptional control by developmental cell signaling. Genes Dev 2002, 16:1167-1181.
- 2. Gerhart J: 1998 Warkany lecture: signaling pathways in development. Teratology 1999, 60:226-239
- Pires-daSilva A, Sommer RJ: The evolution of signalling 3. pathways in animal development. Nat Rev Genet 2003, 4:39-49.
- 4. Cummings FW: On the origin of pattern and form in early Metazoans. Int J Dev Biol 2006, 50:193-208.

Kirschner M, Gerhart J: The plausibility of life: great leaps of 5. evolution. New Haven, CT: Yale University Press; 2005. In this excellent text, the authors provide a thought-provoking view of how

novel biological functions evolve. Bray SJ: Notch signalling: a simple pathway becomes 6

complex. Nat Rev Mol Cell Biol 2006, 7:678-689 The author provides a comprehensive review of Notch signaling.

- Louvi A. Artavanis-Tsakonas S: Notch signalling in vertebrate 7. neural development. Nat Rev Neurosci 2006, 7:93-102.
- Miele L: Notch signaling. Clin Cancer Res 2006, 12:1074-1079. 8.

- 9. Weinmaster G, Kopan R: A garden of Notch-ly delights. Development 2006, 133:3277-3282
- 10. Ehebauer M, Hayward P, Arias AM: Notch, a universal arbiter of cell fate decisions. Science 2006. 314:1414-1415.
- 11. Kopan R: Notch: a membrane-bound transcription factor. J Cell Sci 2002, 115:1095-1097.
- 12. Sundaram MV: The love-hate relationship between Ras and Notch, Genes Dev 2005, 19:1825-1839
- 13. Sundaram MV: Vulval development: the battle between Ras and Notch. Curr Biol 2004. 14:B311-B313.
- 14. Voas MG, Rebay I: Signal integration during development: insights from the Drosophila eye. Dev Dyn 2004, 229:162-175.
- 15. Doroquez DB, Rebay I: Signal integration during development: mechanisms of EGFR and notch pathway function and cross-talk. Crit Rev Biochem Mol Biol 2006, 41:339-385.
- 16. Culi J, Martin-Blanco E, Modolell J: The EGF receptor and N signalling pathways act antagonistically in Drosophila mesothorax bristle patterning. Development 2001, 128:299-308
- 17. zur Lage P, Jarman AP: Antagonism of EGFR and Notch signalling in the reiterative recruitment of Drosophila adult chordotonal sense organ precursors. Development 1999, 126:3149-3157.
- 18. Carmena A, Buff E, Halfon MS, Gisselbrecht S, Jimenez F, Baylies MK, Michelson AM: Reciprocal regulatory interactions between the Notch and Ras signaling pathways in the Drosophila embryonic mesoderm. Dev Biol 2002, 244:226-242.
- 19. Price JV, Savenye ED, Lum D, Breitkreutz A: Dominant enhancers of Egfr in Drosophila melanogaster: genetic links between the Notch and Egfr signaling pathways. Genetics 1997, 147:1139-1153
- 20. Deregowski V, Gazzerro E, Priest L, Rydziel S, Canalis E: Notch 1 overexpression inhibits osteoblastogenesis by suppressing Wnt/β-catenin but not bone morphogenetic protein signaling. J Biol Chem 2006, 281:6203-6210.

In this study, the authors show that Notch/Wnt cross-talk is involved in osteoblastogenesis. Overexpression of NICD is shown to prevent BMP-2 and Wnt effects by suppressing Wnt but not BMP signaling via a mechanism that is likely to be dependent upon the Notch target HES-1.

- 21. Estrach S, Ambler CA, Lo Celso CL, Hozumi K, Watt FM: Jagged 1 is a β -catenin target gene required for ectopic hair follicle formation in adult epidermis. Development 2006, 133:4427-4438.
- 22. Nobta M, Tsukazaki T, Shibata Y, Xin C, Moriishi T, Sakano S, Shindo H, Yamaguchi A: Critical regulation of bone morphogenetic protein-induced osteoblastic differentiation by Delta1/Jagged1-activated Notch1 signaling. J Biol Chem 2005, 280:15842-15848.
- 23. Dahlqvist C, Blokzijl A, Chapman G, Falk A, Dannaeus K, Ibanez CF, Lendahl U: Functional Notch signaling is required for BMP4-induced inhibition of myogenic differentiation. Development 2003, 130:6089-6099.
- 24. Reynolds-Kenneally J, Mlodzik M: Notch signaling controls proliferation through cell-autonomous and non-autonomous

mechanisms in the Drosophila eye. Dev Biol 2005, 285:38-48. In [24°-26°], the authors show that Notch-regulated cell growth during Drosophila eye development involves cross-talk with the Jak/STAT pathway through regulation of the Jak/STAT secreted ligand unpaired.

Chao JL, Tsai YC, Chiu SJ, Sun YH: Localized Notch signal acts 25 through eyg and upd to promote global growth in Drosophila eye. Development 2004, 131:3839-3847.

See annotation to [24*].

- 26. Moberg KH, Schelble S, Burdick SK, Hariharan IK: Mutations in erupted, the Drosophila ortholog of mammalian tumor
- susceptibility gene 101, elicit non-cell-autonomous overgrowth. Dev Cell 2005, 9:699-710. See annotation to [24*].
- 27. Giraldez AJ, Cohen SM: Wingless and Notch signaling provide cell survival cues and control cell proliferation

during wing development. Development 2003, 130:6533-6543.

- Johnston LA, Edgar BA: Wingless and Notch regulate cell-cycle arrest in the developing *Drosophila* wing. *Nature* 1998, 394:82-84.
- 29. van Es JH, van Gijn ME, Riccio O, van den Born M, Vooijs M, • Begthel H, Cozijnsen M, Robine S, Winton DJ, Radtke F *et al.*:
- Begthel H, Cozijnsen M, Robine S, Winton DJ, Radtke F et al.: Notch/γ-secretase inhibition turns proliferative cells in intestinal crypts and adenomas into goblet cells. Nature 2005, 435:959-963.

The authors show that maintenance of undifferentiated, proliferative cells in intestinal crypts requires activation of Notch and Wnt signaling.

- Crosnier C, Stamataki D, Lewis J: Organizing cell renewal in the intestine: stem cells, signals and combinatorial control. Nat Rev Genet 2006, 7:349-359.
- Yu S-Y, Yoo SJ, Yang L, Zapata C, Srinivasan A, Hay BA, Baker NE: A pathway of signals regulating effector and initiator caspases in the developing *Drosophila* eye. *Development* 2002, 129:3269-3278.
- 32. Cordero J, Jassim O, Bao S, Cagan R: A role for wingless in an early pupal cell death event that contributes to patterning the *Drosophila* eye. *Mech Dev* 2004, **121**:1523.
- Mimeault M, Batra SK: Concise review: recent advances on the significance of stem cells in tissue regeneration and cancer therapies. Stem Cells 2006, 24:2319-2345.
- de Lau W, Barker N, Clevers H: WNT signaling in the normal intestine and colorectal cancer. *Front Biosci* 2007, 12:471-491.
- 35. Duncan AW, Rattis FM, DiMascio LN, Congdon KL, Pazianos G,
 Zhao C, Yoon K, Cook JM, Willert K, Gaiano N *et al.*: Integration of Notch and Wnt signaling in hematopoietic stem cell maintenance. Nat Immunol 2005, 6:314-322.

The authors show that Notch is required for Wnt-mediated maintenance of undifferentiated murine hematopoietic stem cells but not for their Wnt-mediated survival or proliferation.

- 36. Androutsellis-Theotokis A, Leker RR, Soldner F, Hoeppner DJ,
- Ravin R, Poser SW, Rueger MA, Bae SK, Kittappa R, McKay RD: Notch signalling regulates stem cell numbers *in vitro* and *in vivo*. *Nature* 2006, **442**:823-826.

The authors identify a general mechanism whereby Notch maintains stem cells through cross-talk with the pI3K/Akt/mTOR and Shh pathways.

Ward EJ, Shcherbata HR, Reynolds SH, Fischer KA, Hatfield SD:
Ruohola-Baker H. Stem cells signal to the niche through the Notch pathway in the *Drosophila* ovary. *Curr Biol* 2006, in press.

In this study, the authors show that, in adult *Drosophila*, germline stem cells (GSCs) require interplay between Notch and TGF- β signals, both for GSC maintenance and generation of the stem cell niche.

- Liu S, Dontu G, Wicha MS: Mammary stem cells, self-renewal pathways, and carcinogenesis. Breast Cancer Res 2005, 7:86-95.
- 39. Allenspach EJ, Maillard I, Aster JC, Pear WS: Notch signaling in cancer. Cancer Biol Ther 2002, 1:466-476.
- Stylianou S, Clarke RB, Brennan K: Aberrant activation of notch signaling in human breast cancer. Cancer Res 2006, 66:1517-1525.
- 41. Dunn KL, Espino PS, Drobic B, He S, Davie JR: **The Ras-MAPK** signal transduction pathway, cancer and chromatin remodeling. *Biochem Cell Biol* 2005, **83**:1-14.
- Derynck R, Akhurst RJ, Balmain A: TGF-β signaling in tumor suppression and cancer progression. *Nat Genet* 2001, 29:117-129.
- 43. Reya T, Clevers H: Wnt signalling in stem cells and cancer. *Nature* 2005, **434**:843-850.
- Pasca di Magliano M, Hebrok M: Hedgehog signalling in cancer formation and maintenance. Nat Rev Cancer 2003, 3:903-911.
- Boudny V, Kovarik J: JAK/STAT signaling pathways and cancer. Janus kinases/signal transducers and activators of transcription. *Neoplasma* 2002, 49:349-355.

- Anzick SL, Kononen J, Walker RL, Azorsa DO, Tanner MM, Guan XY, Sauter G, Kallioniemi OP, Trent JM, Meltzer PS: AIB1, a steroid receptor coactivator amplified in breast and ovarian cancer. Science 1997, 277:965-968.
- 47. Hornberg JJ, Bruggeman FJ, Westerhoff HV, Lankelma J: Cancer: a Systems Biology disease. *Biosystems* 2006, 83:81-90.
- 48. Logan CY, Nusse R: **The Wnt signaling pathway in development** and disease. Annu Rev Cell Dev Biol 2004, **20**:781-810.
- 49. Nusse R: Wht signaling in disease and in development. *Cell Res* 2005, **15**:28-32.
- Ellisen LW, Bird J, West DC, Soreng AL, Reynolds TC, Smith SD, Sklar J: TAN-1, the human homolog of the *Drosophila* notch gene, is broken by chromosomal translocations in T lymphoblastic neoplasms. *Cell* 1991, 66:649-661.
- Weng AP, Ferrando AA, Lee W, Morris JP, Silverman LB, Sanchez-Irizarry C, Blacklow SC, Look AT, Aster JC: Activating mutations of NOTCH1 in human T cell acute lymphoblastic leukemia. Science 2004, 306:269-271.
- Veeraraghavalu K, Subbaiah VK, Srivastava S, Chakrabarti O, Syal R, Krishna S: Complementation of human papillomavirus type 16 E6 and E7 by Jagged1-specific Notch1phosphatidylinositol 3-kinase signaling involves pleiotropic oncogenic functions independent of CBF1;Su(H);Lag-1 activation. J Virol 2005, 79:7889-7898.
- Weijzen S, Rizzo P, Braid M, Vaishnav R, Jonkheer SM, Zlobin A, Osborne BA, Gottipati S, Aster JC, Hahn WC *et al.*: Activation of Notch-1 signaling maintains the neoplastic phenotype in human Ras-transformed cells. *Nat Med* 2002, 8:979-986.
- Fitzgerald K, Harrington A, Leder P: Ras pathway signals are required for notch-mediated oncogenesis. Oncogene 2000, 19:4191-4198.
- Haruki N, Kawaguchi KS, Eichenberger S, Massion PP, Olson S, Gonzalez A, Carbone DP, Dang TP: Dominant-negative Notch3 receptor inhibits mitogen-activated protein kinase pathway and the growth of human lung cancers. *Cancer Res* 2005, 65:3555-3561.
- Stella MC, Trusolino L, Pennacchietti S, Comoglio PM: Negative feedback regulation of Met-dependent invasive growth by Notch. Mol Cell Biol 2005, 25:3982-3996.
- Kiaris H, Politi K, Grimm LM, Szabolcs M, Fisher P, Efstratiadis A, Artavanis-Tsakonas S: Modulation of notch signaling elicits signature tumors and inhibits hras1-induced oncogenesis in the mouse mammary epithelium. *Am J Pathol* 2004, 165:695-705
- Miyamoto Y, Maitra A, Ghosh B, Zechner U, Argani P, Iacobuzio-Donahue CA, Sriuranpong V, Iso T, Meszoely IM, Wolfe MS *et al.*: Notch mediates TGFα-induced changes in epithelial differentiation during pancreatic tumorigenesis. *Cancer Cell* 2003, 3:565-576.
- Hallahan AR, Pritchard JI, Hansen S, Benson M, Stoeck J, Hatton BA, Russell TL, Ellenbogen RG, Bernstein ID, Beachy PA et al.: The SmoA1 mouse model reveals that notch signaling is critical for the growth and survival of sonic hedgehog-induced medulloblastomas. *Cancer Res* 2004, 64:7794-7800.
- 60. Dakubo GD, Mazerolle CJ, Wallace VA: Expression of Notch and Wnt pathway components and activation of Notch signaling in medulloblastomas from heterozygous patched mice. J Neurooncol 2006, **79**:221-227.
- Balint K, Xiao M, Pinnix CC, Soma A, Veres I, Juhasz I, Brown EJ, Capobianco AJ, Herlyn M, Liu ZJ: Activation of Notch1 signaling is required for β-catenin-mediated human primary melanoma progression. J Clin Invest 2005, 115:3166-3176.
- Zagouras P, Stifani S, Blaumueller CM, Carcangiu ML, Artavanis-Tsakonas S: Alterations in Notch signaling in neoplastic lesions of the human cervix. Proc Natl Acad Sci USA 1995, 92:6414-6418.
- 63. Llimargas M: The Notch pathway helps to pattern the tips of the Drosophila tracheal branches by selecting cell fates. Development 1999, **126**:2355-2364.

- 64. Ikeya T, Hayashi S: Interplay of Notch and FGF signaling restricts cell fate and MAPK activation in the *Drosophila* trachea. *Development* 1999, **126**:4455-4463.
- Ghabrial AS, Krasnow MA: Social interactions among epithelial cells during tracheal branching morphogenesis. *Nature* 2006, 441:746-749.
- Steneberg P, Hemphala J, Samakovlis C: Dpp and Notch specify the fusion cell fate in the dorsal branches of the Drosophila trachea. Mech Dev 1999, 87:153-163.
- Chihara T, Hayashi S: Control of tracheal tubulogenesis by Wingless signaling. Development 2000, 127:4433-4442.
- Grishina IB, Kim SY, Ferrara C, Makarenkova HP, Walden PD: BMP7 inhibits branching morphogenesis in the prostate gland and interferes with Notch signaling. *Dev Biol* 2005, 288:334-347.
- 69. Uyttendaele H, Soriano JV, Montesano R, Kitajewski J: Notch4 and Wnt-1 proteins function to regulate branching morphogenesis of mammary epithelial cells in an opposing fashion. *Dev Biol* 1998, **196**:204-217.
- Itoh F, Itoh S, Goumans MJ, Valdimarsdottir G, Iso T, Dotto GP, Hamamori Y, Kedes L, Kato M, ten Dijke Pt P: Synergy and antagonism between Notch and BMP receptor signaling pathways in endothelial cells. *EMBO J* 2004, 23:541-551.
- Chen JY, Bottjer DJ, Oliveri P, Dornbos SQ, Gao F, Ruffins S, Chi H, Li CW, Davidson EH: Small bilaterian fossils from 40 to 55 million years before the Cambrian. *Science* 2004, 305:218-222.
- Nichols SA, Dirks W, Pearse JS, King N: Early evolution of animal cell signaling and adhesion genes. Proc Natl Acad Sci USA 2006, 103:12451-12456.
- Diaz-Benjumea FJ, Cohen SM: Serrate signals through Notch to establish a Wingless-dependent organizer at the dorsal/ ventral compartment boundary of the *Drosophila* wing. *Development* 1995, 121:4215-4225.
- 74. Cho KO, Chern J, Izaddoost S, Choi KW: Novel signaling from the peripodial membrane is essential for eye disc patterning in *Drosophila*. *Cell* 2000, **103**:331-342.
- Galceran J, Sustmann C, Hsu SC, Folberth S, Grosschedl R: LEF1-mediated regulation of Delta-like1 links Wnt and Notch signaling in somitogenesis. *Genes Dev* 2004, 18:2718-2723.
- Tsuda L, Nagaraj R, Zipursky SL, Banerjee U: An EGFR/Ebi/Sno pathway promotes delta expression by inactivating Su(H)/ SMRTER repression during inductive notch signaling. *Cell* 2002, 110:625-637.
- 77. Shaye DD, Greenwald I: LIN-12/Notch trafficking and regulation
 of DSL ligand activity during vulval induction in

Caenorhabditis elegans. Development 2005, **132**:5081-5092. The authors show that crosstalk between Notch and RTK signaling during *C. elegans* vulval development involves an EGFR–Ras–MAPK-mediated promotion of LIN-12/Notch endocytosis and degradation. They identify residues within the LIN-12/Notch intracellular domain important for internalization/degradation and proteins involved in this process.

- Campos LS, Decker L, Taylor V, Skarnes W: Notch, epidermal growth factor receptor, and beta1-integrin pathways are coordinated in neural stem cells. *J Biol Chem* 2006, 281:5300-5309.
- Espinosa L, Ingles-Esteve J, Aguilera C, Bigas A: Phosphorylation by glycogen synthase kinase-3 β down-regulates Notch activity, a link for Notch and Wnt pathways. J Biol Chem 2003, 278:32227-32235.
- Axelrod JD, Matsuno K, Artavanis-Tsakonas S, Perrimon N: Interaction between Wingless and Notch signaling pathways mediated by dishevelled. *Science* 1996, **271**:1826-1832.
- Takizawa T, Ochiai W, Nakashima K, Taga T: Enhanced gene activation by Notch and BMP signaling cross-talk. Nucleic Acids Res 2003, 31:5723-5731.
- 82. Blokziji A, Dahlqvist C, Reissmann E, Falk A, Moliner A, Lendahl U, Ibanez CF: Cross-talk between the Notch and TGF- β signaling

pathways mediated by interaction of the Notch intracellular domain with Smad3. J Cell Biol 2003, 163:723-728.

- 83. Stockhausen MT, Sjolund J, Axelson H: Regulation of the Notch target gene Hes-1 by TGF α induced Ras/MAPK signaling in human neuroblastoma cells. *Exp* Cell Res 2005, **310**:218-228.
- Rohrbaugh M, Ramos E, Nguyen D, Price M, Wen Y, Lai ZC: Notch activation of yan expression is antagonized by RTK/ Pointed signaling in the *Drosophila* eye. *Curr Biol* 2002, 12:576-581.
- Flores GV, Duan H, Yan H, Nagaraj R, Fu W, Zou Y, Noll M, Banerjee U: Combinatorial signaling in the specification of unique cell fates. *Cell* 2000, 103:75-85.
- 86. Hayward P, Brennan K, Sanders P, Balayo T, DasGupta R, Perrimon N, Martinez Arias A: Notch modulates Wnt signalling by associating with Armadillo/β-catenin and regulating its transcriptional activity. *Development* 2005, **132**:1819-1830.
- Eagar TN, Tang Q, Wolfe M, He Y, Pear WS, Bluestone JA: Notch 1 signaling regulates peripheral T cell activation. *Immunity* 2004, 20:407-415.
- 88. Yoo AS, Bais C, Greenwald I: Crosstalk between the EGFR and
 LIN-12/Notch pathways in *C. elegans* vulval development. *Science* 2004, 303:663-666.

The authors show that during *C. elegans* vulval patterning, LIN-12 induces the expression of numerous negative regulators of the EGFR–MAPK pathway in addition to LIP-1.

- Berset T, Hoier EF, Battu G, Canevascini S, Hajnal A: Notch inhibition of RAS signaling through MAP kinase phosphatase LIP-1 during *C. elegans* vulval development. *Science* 2001, 291:1055-1058.
- 90. Wesley CS, Saez L: Notch responds differently to Delta and Wingless in cultured *Drosophila* cells. *J Biol Chem* 2000, 275:9099-9101.
- Masuda S, Kumano K, Shimizu K, Imai Y, Kurokawa M, Ogawa S, Miyagishi M, Taira K, Hirai H, Chiba S: Notch1 oncoprotein antagonizes TGF-beta/Smad-mediated cell growth suppression via sequestration of coactivator p300. *Cancer Sci* 2005, 96:274-282.
- Oswald F, Tauber B, Dobner T, Bourteele S, Kostezka U, Adler G, Liptay S, Schmid RM: p300 acts as a transcriptional coactivator for mammalian Notch-1. *Mol Cell Biol* 2001, 21:7761-7774.
- 93. Hasson P, Egoz N, Winkler C, Volohonsky G, Jia S, Dinur T, Volk T,
- Courey AJ, Paroush Z: EGFR signaling attenuates Grouchodependent repression to antagonize Notch transcriptional output. Nat Genet 2005, 37:101-105.

In this study, the authors provide a potential mechanism for cross-talk between RTK and other pathways with downstream effectors that require the global co-repressor Groucho, such as Notch effectors of the Enhancer of Split family. The authors show that RTK-signaling-responsive phosphorylation of Groucho weakens its capacity to repress, resulting in an attenuation of transcriptional silencing by Enhancer of Split proteins.

- Belandia B, Powell SM, Garcia-Pedrero JM, Walker MM, Bevan CL, Parker MG: Hey1, a mediator of notch signaling, is an androgen receptor corepressor. *Mol Cell Biol* 2005, 25:1425-1436.
- 95. Lawson ND, Vogel AM, Weinstein BM: sonic hedgehog and vascular endothelial growth factor act upstream of the Notch pathway during arterial endothelial differentiation. *Dev Cell* 2002, **3**:127-136.
- Josten F, Fuss B, Feix M, Meissner T, Hoch M: Cooperation of JAK/STAT and Notch signaling in the Drosophila foregut. Dev Biol 2004, 267:181-189.
- McGregor JR, Xi R, Harrison DA: JAK signaling is somatically required for follicle cell differentiation in *Drosophila*. *Development* 2002, 129:705-717.
- Torres IL, Lopez-Schier H, St Johnston D: A Notch/ Delta-dependent relay mechanism establishes anteriorposterior polarity in *Drosophila*. *Dev Cell* 2003, 5:547-558.
- 99. Althauser C, Jordan KC, Deng WM, Ruohola-Baker H: Fringe-dependent notch activation and tramtrack function are

required for specification of the polar cells in Drosophila oogenesis. Dev Dyn 2005, 232:1013-1020.

100. Kamakura S, Oishi K, Yoshimatsu T, Nakafuku M, Masuyama N,

 Gotoh Y: Hes binding to STAT3 mediates crosstalk between Notch and JAK-STAT signalling. Nat Cell Biol 2004, 6:547-554.
 In this paper, the authors show Notch signaling leads to activation of STAT3 through the HES-family-mediated recruitment of JAK2 to STAT3, which leads to STAT3 phosphorylation and activation.

- 101. Kumar JP, Moses K: Egf receptor and notch signaling act upstream of eyeless/pax6 to control eye specification. Cell 2001, 104:687-697.
- 102. Johannes B, Preiss A: Wing vein formation in Drosophila melanogaster: Hairless is involved in the cross-talk between Notch and EGF signaling pathways. Mech Dev 2002, 115:3-14.
- 103. Anderson AC. Kitchens EA. Chan SW. St Hill C. Jan YN. Zhong W. Robey EA: The Notch regulator Numb links the Notch and TCR signaling pathways. J Immunol 2005, 174:890-897.
- 104. Zeng Q, Li S, Chepeha DB, Giordano TJ, Li J, Zhang H, Polverini PJ, Nor J, Kitajewski J, Wang CY: Crosstalk between tumor and endothelial cells promotes tumor angiogenesis by MAPK activation of Notch signaling. Cancer Cell 2005, 8:13-23.
- 105. Dequeant ML, Glynn E, Gaudenz K, Wahl M, Chen J, Mushegian A, Pourquie O: A complex oscillating network of signaling genes underlies the mouse segmentation clock. Science 2006 in press
- 106. Akai J, Halley PA, Storey KG: FGF-dependent Notch signaling maintains the spinal cord stem zone. Genes Dev 2005, 19:2877-2887.
- 107. Yoon K, Nery S, Rutlin ML, Radtke F, Fishell G, Gaiano N: Fibroblast growth factor receptor signaling promotes radial

glial identity and interacts with Notch1 signaling in telencephalic progenitors. J Neurosci 2004, 24:9497-9506.

- 108. Timmerman LA, Grego-Bessa J, Raya A, Bertran E, Perez-Pomares JM, Diez J, Aranda S, Palomo S, McCormick F, Izpisua-Belmonte JC *et al.*: **Notch promotes** epithelial-mesenchymal transition during cardiac development and oncogenic transformation. Genes Dev 2004, 18:99-115.
- 109. Neumann CJ, Cohen SM: A hierarchy of cross-regulation involving Notch, wingless, vestigial and cut organizes the dorsal/ventral axis of the Drosophila wing. Development 1996, 122:3477-3485.
- 110. Couso JP, Martinez Arias A: Notch is required for wingless signaling in the epidermis of Drosophila. Cell 1994, 79:259-272.
- 111. No M. Diaz-Beniumea FJ. Vincent JP. Wu J. Cohen SM: Specification of the wing by localized expression of wingless protein. Nature 1996, 381:316-318.
- 112. Hing HK, Sun X, Artavanis-Tsakonas S: Modulation of wingless signaling by Notch in Drosophila. Mech Dev 1994, 47:261-268.
- 113. de Celis JF, Bray S: Feed-back mechanisms affecting Notch activation at the dorsoventral boundary in the Drosophila wing. Development 1997, 124:3241-3251
- 114. Klein T, Arias AM: Different spatial and temporal interactions between Notch, wingless, and vestigial specify proximal and distal pattern elements of the wing in Drosophila. Dev Biol 1998, 194:196-212.
- 115. Doherty D, Feger G, Younger-Shepherd S, Jan LY, Jan YN: Delta is a ventral to dorsal signal complementary to Serrate, another Notch ligand, in Drosophila wing formation. Genes Dev 1996, 10:421-434.